

## Various Animal Peripheral Platelet Separator Kits

V02

**Size:** 3X200 mL/kit

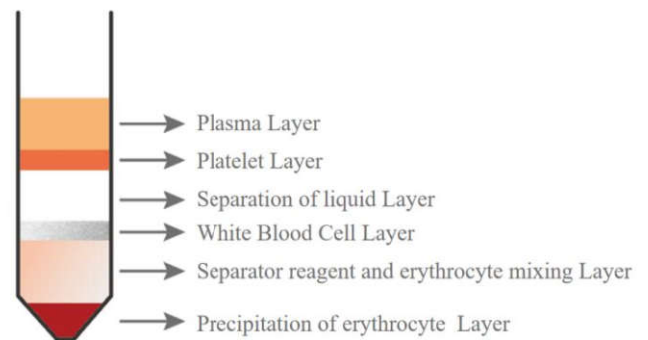
**Storage:** This product is sensitive to light, should avoid light storage at room temperature, shelf life of 2years. After sterile opening, save at room temperature.

### Kit compositions

Kit components	Specifications
Platelet Separation Fluid	200mL
Whole Blood and Tissue Thinning Fluid	200mL
Cell washing solution	200mL

### Protocols(only for reference)

1. Add a minimum of 5mL of separation fluid to the centrifugal tube. If the sample volume is above or equal to 3mL, double the amount of separation fluid. However, make sure that the combined volume of the two fluids does not exceed two-thirds of the centrifugal tube, otherwise it will impact the separation function.
2. First add 4mL of separation solution into the centrifuge tube, then add 2mL of 60% separation solution (1200 $\mu$ L of separation solution +800 $\mu$ L of tissue diluent) to form the separation interface, and finally add the diluted anticoagulant whole blood (note that the total volume of the three can not exceed two-thirds of the centrifuge tube, otherwise it will affect the separation effect).
3. At room temperature, the horizontal rotor 250~350g, centrifugation for 20min(the larger the blood volume, the larger the centrifugal force required, the longer the centrifugation time, the best separation conditions need to be explored).
4. After centrifugation, the centrifuge tube is divided into 6 layers from top to bottom (as shown in the right picture), carefully absorb the platelet and plasma layer into the 15mL clean centrifuge tube, add 10mLPBS or cell washing solution, 500g, centrifugation for 20min.
5. Carefully absorb platelet-rich plasma layers into 15mL clean centrifugal tubes and add 10mL PBS or cell washing fluid (If you need to get platelet-rich plasma, can be absorbed directly, no longer wash) .500g, centrifugal 20minutes.
6. Discard the supernatant, resuspended as single cells.



Separation schematic

### Note

- A. Before opening and mixing upside down with this sterile product, please unseal it under sterile conditions and avoid storing it upside down to extend its storage time.
- B. The separation fluid should always be kept at room temperature (18 $^{\circ}$ C ~ 25 $^{\circ}$ C), if the temperature is low, the separation liquid can be preheated. 4 $^{\circ}$ C or low temperature condition centrifugal, the separation effect is poor.
- C. Inhaling too much platelet plasma in the lower layer of the solution can cause white blood cells at the junction to mix.
- D. Blood samples are best used for fresh anticoagulant blood (within 2h) to avoid freezing and refrigeration.
- E. Use buffers that do not contain Ca, Mg, or Mn ions when diluting blood or washing cells, as they can decrease cell yield and purity.
- F. The separation effect can be influenced by the viscosity of blood samples or differences in temperature. Regulating the number of centrifugal rotations and centrifugal time can lead to the best separation conditions.
- G. When developing a separation fluid, users should mention the cell dispersion coefficient and cell charge of different animal blood in different specific weight separation fluids.



### Related products

- YA0902 *Disposable Pasteurized Straw*
- R1018 *Cell Wash Solution*
- R1017 *Whole Blood and Tissue Diluent*
- S9020 *Superior Fetal Bovine Serum*
- T1300 *Trypsin-EDTA Digest (0.25%) Contains no Phenol Red*  
*A Variety of Other Animal and Other Cell Separations and Kits*

Note: For more literature on the use of this product, please refer to Solarbio's official website.

