

## Total Protein Sulphydryl Content Assay Kit

**Note:** Take two or three different samples for prediction before test.

**Operation Equipment:** Spectrophotometer

**Catalog Number:** BC5800

**Size:** 50T/48S

### Components:

**Extract I:** Liquid 40 mL ×1. Storage at 2-8°C.

**Extract II:** Liquid 40 mL ×1. Storage at 2-8°C.

**Preparation of extract solution:** before use according to the sample volume in accordance with the extract I: extract II = 1mL: 1 mL for the preparation, ready to use, do not mix all at once.

**Reagent I:** Liquid 125 mL ×1. Storage at 2-8°C.

**Reagent II:** Liquid 20 mL ×1. Storage at 2-8°C. If reagent II precipitated, reagent two can be placed in a 37 °C water bath heating until clarified and transparent after use.

**Reagent III:** Liquid 18 mL ×1. Storage at 2-8°C.

**Reagent IV:** Liquid 3 mL ×1. Storage at 2-8°C.

**Powder I:** Powder ×1. Storage at 2-8°C.

**Standard:** Powder ×1, Store at 2-8°C. 10 mg reduced glutathione (GSH). It was prepared to 25 μmol/mL by adding 1.3 mL of distilled water before use and can be stored at 2-8°C for 4 weeks.

**0.125μmol/mL standard preparation:** take 50μL of 25μmol/mL standard, add 950μL of distilled water, mix thoroughly to formulate 1.25μmol/mL standard; then take 100μL of 1.25μmol/mL standard, add 900μL of distilled water, mix thoroughly to formulate 0.125μmol/mL standard.

### Product Description

The presence of sulphydryl allows proteins to undergo disulfide bond formation, thereby maintaining molecular stability and functionality. Sulphydryl are also involved in redox reactions and have important biological roles. In cells, changes in sulphydryl content are closely related to the onset and progression of a variety of diseases, and thus sulphydryl have become an important object of study in the biomedical field. **This kit determines the sum of sulphydryl produced by protein disulfide bond breakage and its own free sulphydryl .**

Reducing agent will cause disulfide bond cleavage to generate sulphydryl , sulphydryl will undergo nucleophilic reaction, that is, sulphydryl reacts with 5,5'-dithio-bis-nitrobenzoic acid (DTNB) to produce a yellow color compound, with a maximum absorption peak at 412 nm, from which the total sulphydryl content of proteins can be calculated.

### Reagents and Equipment Required but Not Provided.

Spectrophotometer, tabletop centrifuge, water bath, adjustable pipette, 1 mL glass cuvette, Acetone, mortar/homogenizer/ sonicator and distilled water.

## Procedure

### I. Sample Processing

1. Tissue: according to the ratio of mass (g): volume of extract (mL) is 1:5~10 (it is recommended to weigh about 0.1g, add 1mL of extract) add extract, homogenize in ice bath and centrifuge at 4°C, 3000rpm for 10min, discard the supernatant. Add 2mL of Reagent I to the precipitate, stir well to dissolve the precipitate, and use the precipitate solution as a sample for the experiment. (Note: (1) plant leaves and other samples with high fiber content, dissolve the precipitate and centrifuge at 4°C and 3000rpm for 3min, then take the supernatant as the sample for the experiment; (2) a lot of air bubbles will be generated after adding Reagent I, please add slowly, and it is recommended to use 5mL EP tubes.)

2. Bacteria/cells: according to the ratio of the number of bacteria/cells ( $10^6$ ): the volume of extraction solution (mL) is 5~10:1 (it is recommended that 5 million bacteria/cells added to 1mL of the extraction solution), ultrasonic crushing in an ice bath (power of 200W, ultrasound for 3 seconds, an interval of 10 seconds, a total of 3min), centrifuged at 4°C, 3,000 rpm for 10min, and discarded the supernatant. Add 2mL of Reagent I to the precipitate, stir well to dissolve the precipitate, and use the precipitate solution as a sample for the experiment. (Note: (1) If the precipitate is not completely dissolved, centrifuge at 4°C and 3000rpm for 3min, and take the supernatant as sample for experiment; (2) a lot of air bubbles will be generated after adding Reagent I, please add it slowly, and it is recommended to use a 5mL EP tube).

3. Serum/plasma, milk and other liquids: Take 100 $\mu$ L of liquid sample and add 0.9mL of acetone, centrifuge at 4°C, 3000rpm for 10min, discard the supernatant. Add 2mL of Reagent I to the precipitate, stir well to dissolve the precipitate, and the precipitate dissolved solution is used as the sample for the experiment. (Note: If the measured value is small, you can change the ratio of sample to acetone, such as taking 0.2mL liquid sample and adding 0.8mL acetone or 0.3mL liquid sample and adding 0.7mL acetone, pay attention to synchronous modification of the calculation formula).

### II. Determination Procedure

1. Preheat the spectrophotometer for more than 30 minutes, adjust the wavelength to 412 nm and set the counter to zero with distilled water.

2. Operation table (recommended for operation in 5mL EP tubes)

Reagent Name (mL)	Test Tube ( $A_T$ )	Blank Tube ( $A_B$ )	Standard Tube ( $A_S$ )
Sample	0.5	-	-
Distilled Water	-	0.5	-
Standard	-	-	0.5
Powder I	5 mg	-	-
React for 30min with the lid open, during the period every 10min with the pipette tip blowing until the bubble is no longer produced, it is prohibited to withhold the lid.		-	-

extract solution	0.3	0.3	0.3
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Slowly add the extract and mix well, and blow repeatedly with a pipette tip until bubbles are no longer produced (during which time a large number of bubbles will be produced and left uncapped)		-	-
Reagent II	0.3	0.3	0.3
The supernatant was centrifuged at 3,000 rpm for 10 min at 4°C and then placed in a 1.5 mL EP tube.		-	-
Supernatant	0.7	0.7	0.7
Reagent III	0.25	0.25	0.25
The absorbance at 412 nm was measured and recorded as <b>Ac</b> .		-	-
Reagent IV	0.05	0.05	0.05
Mix well, let it stand at room temperature for 10 min and then determine the absorbance at 412 nm, which was recorded as $A_T$ , $A_C$ , $A_B$ and $A_S$ , respectively. Calculate $\Delta A_T = A_T - A_C$ , $\Delta A_S = A_S - A_B$ . The blank and standard tubes should only be measured 1-2 times. Note: In the first step of determining the absorbance at 412 nm, all the reaction solution can be poured into a 1 mL glass cuvette for determination, after which the reagent can be added directly to the cuvette IV mixing and then continue the determination.			

### III. Calculation Formula

#### 1. Calculate

##### 1) Calculate by protein concentration

$$\begin{aligned} \text{Total Protein Sulphydryl content } (\mu\text{mol/mL prot}) &= \Delta A_T \div (\Delta A_S \div C_s) \times V_s \div (V_s \times C_{pr}) \times F \\ &= 0.125 \times \Delta A_T \div \Delta A_S \times C_{pr} \times F \end{aligned}$$

##### 2) Calculate by sample weight

$$\begin{aligned} \text{Total Protein Sulphydryl content } (\mu\text{mol/g weight}) &= \Delta A_T \div (\Delta A_S \div C_s) \times V_r \div W \times F \\ &= 0.25 \times \Delta A_T \div \Delta A_S \div W \times F \end{aligned}$$

##### 3) Calculate by the Liquid volume

$$\text{Total Protein Sulphydryl content } (\mu\text{mol/mL}) = \Delta A_T \div (\Delta A_S \div C_s) \times V_r \div V_s \times F = 2.5 \times \Delta A_T \div \Delta A_S \times F$$

##### 4) Calculate by the number of cells

$$\begin{aligned} \text{Total Protein Sulphydryl content } (\mu\text{mol}/10^6 \text{ cell}) &= \Delta A_T \div (\Delta A_S \div C_s) \\ \times V_r \div N \times F &= 0.25 \times \Delta A_T \div \Delta A_S \div N \times F \end{aligned}$$

$C_s$ : Standard Tube Concentration, 0.125 μmol/mL;  $V_s$ : Volume of sample added, 0.5 mL;  $C_{pr}$ : Sample Protein Concentration, mg/mL, Protein concentration is measured separately. The BCA method is recommended.;  $W$ : sample quality, g;  $V_r$ : Volume of reagent I added during extraction,

2mL; Vsl: Sample volume of liquid added during extraction, 0.1mL; F: dilution factor; N: Total number of cells/bacteria, in  $10^6$ .

**Note:**

1. If the  $\Delta A$  of the sample is  $<0.01$ , the sample volume can be increased appropriately and then measured, paying attention to the simultaneous modification of the blank and standard tubes and the calculation formula; if the  $\Delta A$  of the sample is  $>1.5$ , the precipitation solution can be diluted with reagent I and then measured, paying attention to the simultaneous modification of the dilution factor in the calculation formula.

**Examples:**

1. Take 100 $\mu$ L horse serum, according to the assay procedure, with 1mL glass cuvette measured  $\Delta A_T = A_T - A_c = 0.557 - 0.038 = 0.519$ ,  $\Delta A_S = A_S - A_B = 0.633 - 0.076 = 0.557$ , according to the sample mass calculation of the total sulfhydryl content of proteins obtained:

$$\text{Total protein sulfhydryl content } (\mu\text{mol/mL}) = 2.5 \times \Delta A_T \div \Delta A_S = 2.329 \mu\text{mol/mL}.$$

2. 0.1036g of mouse liver was taken and operated in accordance with the assay steps.  $\Delta A_T = A_T - A_c = 0.727 - 0.111 = 0.616$ ,  $\Delta A_S = A_S - A_B = 0.633 - 0.076 = 0.557$ , and the total sulfhydryl content of protein was calculated according to the sample mass:

Total protein sulfhydryl content ( $\mu\text{mol/g weight}$ ) =  $0.25 \times \Delta A_T \div \Delta A_S \div W \times F = 2.669 \mu\text{mol/g weight}$ .

3, take 0.1078g of soybean powder, precipitation solution diluted 2 times with reagent I, in accordance with the measurement steps, using a 1mL glass cuvette measured  $\Delta A_T = A_T - A_c = 1.080 - 0.068 = 1.012$ ,  $\Delta A_S = A_S - A_B = 0.633 - 0.076 = 0.557$ , according to the mass of the sample calculation of the total sulfhydryl content of protein to get:

Total protein sulfhydryl content ( $\mu\text{mol/g weight}$ ) =  $0.25 \times \Delta A_T \div \Delta A_S \div W \times F = 8.427 \mu\text{mol/g weight}$

**Related Products:**

- BC1370/BC1375 Total Mercapto(-SH) Content Assay Kit
- BC1430/BC1435 Thiol Content Assay Kit (Non-Protein Sample)
- BC5800/BC5805 Total Protein Thiol Content Assay Kit
- BC5890/BC5895 Protein Free Thiol Content Assay Kit